ARTICLE

www.rsc.org/obc

Synthesis and properties of bioactive 2- and 3-amino-8-methyl-8Hquino[4,3,2-kl]acridine and 8,13-dimethyl-8H-quino[4,3,2-kl]acridinium salts †

Ian Hutchinson, Andrew J. McCarroll, Robert A. Heald and Malcolm F. G. Stevens* Cancer Research Laboratories, School of Pharmacy, University of Nottingham, Nottingham, UKNG72RD

Received 4th September 2003, Accepted 7th November 2003 First published as an Advance Article on the web 10th December 2003

Cyclisation of 9-(benzotriazol-1-yl)acridine 1 to the pentacycle 8H-quino[4,3,2-kl]acridine 5 in a range of low-boiling solvents is mechanistically distinct from previously published photochemical (carbene) and thermolytic (radical) cyclisations. Fragmentation of the triazole ring of 1 to a diazonium intermediate 7, and its subsequent heterolysis $(-N_2)$ and cyclisation is facilitated by solvation of intermediate zwitterionic species. Derivatives of 2- and 3-aminoquinoacridines methylated in the 8-position can be converted to 8,13-dimethylquino[4,3,2-kl]acridinium iodide salts with methyl iodide and were required for biological examination as potential telomerase inhibitors. The chloro group in 3-chloro-8-methyl-8H-quino[4,3,2-kl]acridine can be replaced efficiently by benzylamino, 4-morpholinyl and cyano substituents in palladium(0) mediated reactions.

Introduction

9-(Benzotriazol-1-yl)acridine 1 is a convenient precursor for the synthesis of the parent 8H-quino[4,3,2-kl]acridine ring-system 5. Mitchell and Rees reported a photochemical conversion of $1 \rightarrow 5$ in acetonitrile and proposed that an intermediate carbene 2 (derived initially from a triplet diradical) in the guise of its "dipolar form" 4 cyclised to the pentacycle 5 (Scheme 1).² We have previously probed the parallel thermal (Graebe-Ullmann)³ transformation $1 \rightarrow 5$ by differential scanning calorimetry.⁴ The coincident melting and exothermic degradation $(-N_2)$ of 1 occurs near explosively at 245.6 °C. The reaction can be controlled on a preparative scale in boiling diphenyl ether (bp 259 °C) and we have proposed that a diradical reactive intermediate 3 is involved in these cases.⁴ The same quinoacridine 5 can be prepared from 9-(2-bromoanilino)acridine 6 with tributyltin hydride–AIBN in boiling toluene in a process that is undoubtedly radical in character.^{4,5}

Surprisingly, a reinvestigation of the thermolysis of 1 exposed a new twist to these mechanistic interpretations: the degradation of 1 in 98% yield in boiling triglyme (216 °C), and

† Part 15 in the series: Antitumour polycyclic acridines. See ref. 1 for part 14.

vields of 98% in ethanol (78 °C) and 95% in methanol (65 °C), cannot be accommodated reasonably within the diradical hypothesis. In this paper we propose a new mechanism to account for the low temperature conversion $1 \rightarrow 5$ and have adopted this route to prepare 2-nitro- and 3-chloro-quino[4,3,2-kl]acridines which can be further processed to a range of more interesting and bioactive pentacyclic quinoacridinium salts. In particular, to further progress our search for potent telomerase inhibitors of the quadruplex DNA-stabilising class,^{1,6,7} we sought methods for making 8,13-dimethylquinoacridinium salts with amino or acylamino substituents in the 2- and 3-positions.

Chemistry

Degradation of 9-(benzotriazol-1-vl)acridine

Results of the thermolysis of 1 in a range of solvents are recorded in Table 1. Whereas thermolysis in boiling diphenyl ether and triglyme was essentially complete and high-yielding in 1 or 2 hours, respectively, no reaction took place after prolonged boiling in diglyme (bp 162 °C). Most unexpected were results obtained with a range of alcohols: although a satisfactory yield of 5 was obtained with the primary alcohol



Solvent	Temperature/°C	Reaction time/h	Yield (%) ^b	
Methanol	65	36	95	
Ethanol	78	24	98	
Trifluoroethano	1 88	72	0	
Propan-1-ol	97	24	Trace ^c	
Propan-2-ol	82	29	50	
Butan-1-ol	117	24	90	
Butan-2-ol	98	24	Trace ^c	
Diglyme	162	24	0	
Triglyme	216	2	98	
Diphenvl ether	259	1	97	
Benzene	80	48	0	
Toluene	110	48	0	
Triethylamine	89	24	80	
Pvridine	115	24	60	
Dimethylformar	nide 156	24	80	

Reactions conducted on a 0.5 g scale. ^b Determined by ¹H NMR. ^c Identified by TLC.

220



Scheme 1 Reagents and conditions: (i) hv, acetonitrile; (ii) heat, 259 °C; (iii) Bu₃SnH, AIBN, toluene, reflux.

butan-1-ol, only trace amounts of pentacycle **5** were formed from butan-2-ol and propan-1-ol; and no conversion was observed in trifluoroethanol. Contrary to expectations, clean conversion $1 \rightarrow 5$ was evident in the lower boiling alcohols ethanol and methanol. Attempts to catalyse the reaction in alcohols with acids (acetic, TFA, HCl gas) supressed the desired reaction since acridine ring protonation renders the benzotriazolyl residue vulnerable to nucleophilic displacement by the alcohol.⁴ To complete the picture, thermolyses of **1** in the bases triethylamine and pyridine and in DMF also afforded **5** but no conversion was observed in boiling non-polar solvents toluene and benzene (Table 1).

The presence of both acridine and benzotriazolyl moieties are essential to facilitate degradation of 1 since no cyclisation was observed when 9-(1,2,3-triazol-1-yl)acridine or 1-phenyl-1,2,3-benzotriazole were refluxed in low-boiling alcohols. A possible mechanism involves the formation of a diazonium species 7 (Scheme 2) which can cyclise to quinoacridine 5 *via* zwitterion 4 and the carbenium ion reactive species 8, in a process showing affinities to the intramolecular arylation by diazonium compounds (Pschorr cyclisation),⁸ but without the necessity for copper catalysis. A combination of several factors – the propensity of benzotriazole to undergo N–N bond cleavage, resonance stabilisation, solvation of cationic reactive



Scheme 2 Reagents and conditions: solvent (see Table 1), reflux.

species – must all contribute to the initiation of heterolytic fission of the benzotriazole. Because the transformation $1 \rightarrow 5$ does not proceed in boiling acetonitrile alone, this third process is mechanistically distinct from that proposed for the photochemical transformation in acetonitrile by Mitchell and Rees² and also from the diradical (Graebe–Ullmann) process which occurs thermally at 245.6 °C. We admit that our interpretation of this process (Scheme 2) doesn't fully explain the puzzling results in Table 1, particularly the apparent 'all-or-nothing' effect in the range of alcohols studied. Further mechanistic nuances of this reaction may be revealed by a more detailed investigation.[‡]

Synthesis and chemistry of 2-aminoquino[4,3,2-kl]acridine

9-(6-Nitrobenzotriazol-1-yl)acridine **9**, prepared by nitrosation of 9-(2-amino-5-nitroanilino)acridine,⁴ was thermolysed most efficiently in boiling triglyme in which it is soluble; the product, 2-nitroquinoacridine **10** (95%), was precipitated with water. Because of the insolubility of **9** in boiling ethanol or methanol, formation of **10** was very slow in these solvents. (This conversion has been effected previously in 65% yield in boiling diphenyl ether.⁴)

Methylation of 10 with NaH-dimethyl sulfate gave the 8-methyl-2-nitroquinoacridine 11 which was reduced to the corresponding amine 12 with stannous chloride in concentrated hydrochloric acid. Acylation of 12 with a range of acid anhydrides or acid chlorides furnished the amides 13a-g which were then processed to the 8,13-dimethylquinoacridinium iodide salts 14a-g with methyl iodide at 150 °C for 2 days (Scheme 3). Attempts to deacetylate 14a in aqueous sodium hydroxide led to demethylation at the 13-position and the isolation of minor yields of the amide 13a and its precursor amine 12.

The structure of amine 12 was confirmed using COSY and NOESY 2D NMR. The proton at δ 6.99 shows only a small *meta*-coupling (2.2 Hz) with the proton at 6.88, indicating that

[‡] We thank Professor C. W. Rees for helpful discussion on this mechanism.



Scheme 3 Reagents and conditions: (i) Triglyme, 216 °C, 2 h, then H₂O; (ii) NaH, Me₂SO₄, in DMF, 25 °C, 1 h; (iii) SnCl₂·2H₂O, 10 M HCl, 25 °C, 5 d; (iv) R¹COCl, NEt₃, DCM, cat DMAP, 0 °C; or (R¹CO)₂O, pyridine, reflux, 1 h; (v) NaNO₂, 2 M HCl; (vi) MeI, 150 °C, 2 d; (vii) KI, 80 °C, 0.5 h.

these protons are in the C-1 and C-3 positions respectively. (For numbering of the quinoacridine ring see structure **5**, Scheme 1.) The COSY spectrum also revealed the three coupling networks present: 6.99–6.88–8.19, 7.87–7.71–7.12, and 7.5*–7.5*–7.22– 8.72. (The asterisks indicate that these protons had signals which overlapped, and were unresolved.) When the proton at δ 8.19 was irradiated in an NOE difference experiment, the protons at 6.88 and 7.87 displayed positive NOEs, hence the assignment of the proton at 7.87 to the 5-position. Protons at δ 7.12 and 7.55 displayed positive NOEs when the N-methyl group at δ 3.66 was irradiated, confirming the assignment of the proton at C-7. These assignments are in good agreement with those of other 8-methylquinoacridines.⁹

Nitrosation of amine **12** gave the diazonium salt **15** which was converted directly to the 2-iodoquinoacridine **16** by a Sandmeyer reaction and subsequently converted to the methiodide salt **17** (Scheme 3).

Synthesis of 2-amino-8,13-dimethylquinoacridinium salts 20 was best approached from 12 through the Boc-protected amine 18 which was methylated under mild conditions (80 °C for 7 days) with methyl iodide to give the protected aminoquino-acridinium salt 19. Deprotection of 19 with aqueous HCl gave the chloride salt 20a after anion exchange; the same product was formed less efficiently by hydrolysis of the acetamide 14a in HCl (Scheme 4). Methylation of 18 under 'normal' conditions (methyl iodide at 150 °C) led to removal of the Boc group and formation of a mixture of the quaternary iodides 20b-d which could be separated, with difficulty, by column chromatography.

Synthesis and chemistry of 3-aminoquino[4,3,2-kl]acridine

The 3-chloro-8-methyl-quinoacridine 23 was identified as a suitable starting precursor for palladium(o) mediated conversion to quinoacridines 25 and 26 bearing nitrogen-containing functionalities in the 3-position. 9-(5-Chlorobenzotriazol-1yl)acridine 21 was thermolysed efficiently in boiling triglyme to furnish the 3-chloroquinoacridine 22 (95%) which was methylated (NaH-dimethyl sulfate) to give 23 (Scheme 5). Alternatively the N-methyl-acridone 24, which has been prepared from 1-bromo-10-methylacridone by a cross-coupling route, can be cyclised to the same quinoacridine 23 in POCl₃.¹ Coupling between 23 and benzylamine was accomplished in the presence of Pd₂(dba)₃ and P(t-Bu)₃ in dioxane. The benzylamine 25a was debenzylated with hydrazine hydrate in the presence of Pd/C catalyst in methanol to yield the 3-aminoquinoacridine 25b which was acetylated in acetic anhydridepyridine to furnish the acetamide 25c. Similar coupling between 23 and morpholine gave the 3-morpholinyl-quinoacridine 25d and cyanation of 23 was effected under Jin-Confalone conditions.^{io} The 3-cyanoquinoacridine **25e** (98% yield) ($v_{max} \subset \mathbb{E}N$, 2215 cm⁻¹) offered the prospect of further conversion to a 3-(aminomethyl)quinoacridine 25f but this reduction could not be effected with LiAlH₄, catalytic hydrogenation over palladium-charcoal, or SmI₂ electron transfer reduction. Finally, acetamide 25c was converted to the quaternary iodide salt 26 with methyl iodide at 150 °C.

Because of the difficulty in gaining reliable CHN microanalysis on these polycyclic acridines which co-crystallise with variable proportions of solvents, a feature of this class of compound,^{1,4} all compounds were characterised by ¹H NMR and HRMS. Prior to biological evaluation compounds were shown to be single entities by TLC.

Biological results

Several 2-substituted quino[4,3,2-*kl*]acridines were tested for growth-inhibitory activity¹¹ against human cancer cells in the US National Cancer Institute NCI *in vitro* assay (Table 2). Results showed, that against 60 cell types, the mean growth-inhibitory activities (mean GI₅₀ values) varied over a >2 log range. The most potent agents were the 8,13-dimethyl-8*H*-quino[4,3,2-*kl*]acridinium iodide salts **20c** and **20d** with GI₅₀ values <1 μ M and selective activities against cells of the colon and melanoma sub-panels (data not shown). However, these 2-day drug exposure assay results disguise other valuable mechanistic information which can be gleaned from the information-rich screen by COMPARE analysis. COMPARE is a computerised pattern-recognition algorithm used to analyse information generated by the NCI screen ¹² and is a method for determining and expressing the degree of similarity, or lack



Scheme 4 Reagents and conditions: (i) Boc anhydride, NaHCO₃, THF; (ii) MeI, 80 °C, 7 d; (iii) MeI, 150 °C, 48 h; (iv) 2 M HCl, MeOH.



Scheme 5 Reagents and conditions: (i) triglyme, 216 °C, 2 h, then H_2O ; (ii) NaH, Me_2SO_4 , in DMF, 25 °C, (iii) see ref. 1; (iv) for 25a, PhCH₂NH₂, Pd₂(dba)₃, NaOtBu, dioxane, 120 °C, 24 h; (v) NH₂NH₂·H₂O, Pd on C, MeOH, 65 °C, 4 h; (vi) Ac₂O, pyridine, 100 °C, 2 h; (vii) MeI, 150 °C, 3 d.

Table 2Growth inhibitory activity of 2-substituted quino[4,3,2-kl]acridines against human cancer cell lines in the National Cancer Institute (USA)60 cell panel^a



thereof, of mean graph profiles generated from structurally similar (or disparate) compounds. Agents with matching response fingerprints and Pearson Correlation Coefficients (PCCs) > 0.6 can be deduced as having similar biological mechanisms; and the higher the PCC value the greater the confidence in this interpretation. Compounds **13a**, **14a**, **20c** and **20d** were used to explore these relationships. Thus when the neutral 2-acetylamino-quinoacridine **13a** was employed as 'seed' (PCC 1.00), cross PCC values of <0.5 indicated that there were no mechanistic similarities to the three other compounds. In contrast, the trimethyl and tetramethyl quaternary salts **20c,d** were strongly mechanistically related to each other (PCC 0.89), but correlation of the profile of the 2-acetylamino-quinoacridinium salt **14a** to those of **20c** and **20d** was weaker (PCC 0.59 and 0.6, respectively).

COMPARE can also be mobilised to relate biological profiles of new investigational agents to those of clinically-used agents of defined mechanism. In this analysis, the methylamines **20c** and **20d** displayed PCC values >0.6 to a range of natural product DNA-binding agents (*e.g.* actinomycin D, rubidazone, adriamycin, daunomycin, bactobolin, deoxydoxorubicin, bruceantin); also the compounds were predicted to be substrates for *P*-glycoprotein-mediated drug efflux. In contrast, the acetylamino-derivative **14a** gave no PCC values > 0.6 to clinically-used agents, suggesting that it operated by a different biological mechanism. Indeed in other (unpublished) work we have shown that **14a** is a potent telomerase inhibitor (IC₅₀ 0.375 μ M) and its relative lack of cytotoxicity (mean GI₅₀ 12 μ M), coupled to a simple 5-step synthesis, has led us to select this agent for further detailed biological and physico-chemical evaluation.

Experimental

Melting points were measured on a Gallenkamp apparatus and are uncorrected. IR spectra were recorded as KBr discs on a Perkin Elmer Spectrum One FT-IR spectrometer. Mass spectra were recorded on either a Micromass Platform spectrometer, an AEI MS-902 (nominal mass), or a VG Micromass 7070E or a Finigan MAT900XLT spectrometer (accurate mass). NMR spectra were recorded on a Bruker ARX 250 instrument at 250.13 MHz (¹H), 62.9 MHz (¹³C) and 235.3 MHz (¹⁹F) in $[^2H_6]$ DMSO or CDCl_3; coupling constants are in Hz. Merck silica gel 60 (40–60 $\mu M)$ was used for column chromatography.

9-(Benzotriazol-1-yl)acridine $1, ^4$ 9-(6-nitrobenzotriazol-1-yl)acridine $9, ^4$ 9-(5-chlorobenzotriazol-1-yl)acridine $21, ^4$ 3-chloro-8*H*-quino[4,3,2-*kl*]acridine 22^4 and 3-chloro-8-methyl-8*H*quino[4,3,2-*kl*]acridine 23^7 were prepared as indicated.

Thermolysis of 9-(benzotriazol-1-yl)acridines

The benzotriazolylacridine **1** was heated in a range of solvents for the indicated time (see Table 1). 9-(6-Nitrobenzotriazol-1yl)acridine **9** was boiled in triglyme for 2 h. 2-Nitro-8*H*-quino-[4,3,2-*kl*]acridine **10** was isolated in 95% yield following precipitation with water. The product had identical IR, ¹H and ¹³C NMR spectra to a previously prepared sample.⁴

8-Methyl-2-nitro-8H-quino[4,3,2-kl]acridine 11

2-Nitro-8*H*-quino[4,3,2-*kl*]acridine (0.5 g, 1.6 mmol) was dissolved in DMF (10 cm³) and added to a suspension of sodium hydride (0.1 g, 4.2 mmol) in DMF (10 cm³). After stirring for 30 minutes at 25 °C, dimethylsulfate (0.3 g, 2.4 mmol) was added dropwise and stirring continued for a further 1 h. The reaction mixture was poured into water (50 cm³). The resulting precipitate was collected by filtration and washed with water (50 cm³) to give a dark red solid (0.5 g, 95%), mp 285 °C (dec.); ν_{max}/cm^{-1} 1595 (C=N); $\delta_{\rm H}$ ([²H₆] DMSO) 8.67 (1H, d, *J* 7.75), 8.61 (1H, d, *J* 9.0), 8.42 (1H, d, *J* 2.5), 8.11–8.05 (2H, m), 7.86 (1H, t, *J* 8.5), 7.68–7.55 (2H, m), 7.45 (1H, d, *J* 8.25), 7.24 (1H, t, *J* 7.0), 3.67 (3H, s, CH₃); *m*/*z* (ES-HRMS) 328.1078 [C₂₀H₁₄N₃O₂⁺ requires 328.1081].

2-Amino-8-methyl-8*H*-quino[4,3,2-*kl*] acridine 12

A mixture of 8-methyl-2-nitro-8*H*-quino[4,3,2-*kl*] acridine (17 g, 52 mmol) and SnCl₂·2H₂O (52 g, 230 mmol) in 10 M HCl (300 cm³) was stirred at room temperature for 60 hours. After this time, 10 M NaOH solution was added until the pH reached 12, then the mixture was filtered and washed with water. Column chromatography (CHCl₃ \rightarrow CHCl₃–MeOH) yielded impure material which was further purified by washing with chloroform, yielding 2-amino-8-methyl-8*H*-quino[4,3,2-*kl*]-

acridine as a brown–red powder (9.43 g; 61%), mp > 250 °C; $\delta_{\rm H}([^2{\rm H}_6]{\rm DMSO})$ 8.72 (1H, dd, *J* 1.3 and 7.9), 8.19 (1H, d, *J* 8.8), 7.87 (1H, d, *J* 8.1), 7.71 (1H, t, *J* 8.1), 7.51–7.63 (2H, m), 7.22 (1H, td, *J* 1.3 and 7.2), 7.12 (1H, d, *J* 8.2), 6.99 (1H, d, *J* 2.2), 6.88 (1H, dd, *J* 2.3 and 8.7), 5.61 (2H, s, NH₂), 3.66 (3H, s, CH₃); $\delta_{\rm C}$ ([²H₆] DMSO) 150.0, 148.6, 146.8, 141.2, 141.2, 134.7, 131.9, 131.6, 124.9, 123.7, 121.5, 120.9, 115.3, 114.6, 114.3, 113.2, 110.0, 108.5, 106.3, 33.4; *m/z* (EI-HRMS) 297.1255 [C₂₀H₁₅N₃⁺ requires 297.1266].

Acylation of 2-amino-8-methyl-8H-quino[4,3,2-kl]acridine 12

General method A

To a solution of amine **12** (1.0 mmol) in DCM (5 cm³) at 0 °C was added acid chloride (1.0 mmol), triethylamine (1.0 mmol), and catalytic DMAP. The mixture was stirred overnight, then evaporated to dryness. Water (20 cm³) was added, and the solid precipitate was collected and purified by column chromatography (5–10% MeOH–CHCl₃). Alternatively, when using an acid anhydride, the reaction was carried out in refluxing pyridine for 1 h. The following acylamides were prepared:

2-Acetylamino-8-methyl-8H-quino[4,3,2-kl]acridine 13a.

Method A, from **12** and acetic anhydride (2.0 mol. equiv.) in pyridine, acetylamino-quinoacridine **13a** was isolated as a tan solid (79%), mp 340–345 °C; v_{max}/cm^{-1} 3250 (NH), 1657 (C=O); $\delta_{\rm H}$ ([²H₆]DMSO) 10.25 (1H, br s, NH), 8.75 (1H, dd, *J* 1.5 and 8.0), 8.42 (1H, d, *J* 9.0), 8.31 (1H, d, *J* 1.8), 8.00 (1H, d, *J* 8.0), 7.80 (1H, t, *J* 8.0), 7.57–7.63 (3H, m), 7.23–7.30 (2H, m), 3.68 (3H, s, NCH₃), 2.16 (3H, s, CH₃); *m/z* (ES-HRMS) 340.1445 [C₂₂H₁₇N₃O + H⁺ requires 340.1450].

8-Methyl-2-trifluoroacetylamino-8H-quino[4,3,2-kl]acridine

13b. Method A, from **12** and trifluoroacetic anhydride, the trifluoroacetylamido-quinoacridine **13b** was isolated as an orange powder (87%), mp 230 °C (dec.); v_{max}/cm^{-1} 3280 (NH), 1701 (C=O); $\delta_{F}([^{2}H_{6}]DMSO)$ -73.2; m/z (EI-HRMS) 393.1077 [C₂₂H₁₄N₃F₃O⁺ requires 393.1089].

2-n-Butylamido-8-methyl-8H-quino[4,3,2-kl]acridine 13c.

Method A, from **12** and valeryl chloride, the quinoacridine **13c** was isolated as a red powder after column chromatography (78%). Alternatively, the product crystallised slowly from CHCl₃–MeOH and was filtered and washed with chloroform, giving yellow plates, mp 272–273 °C; ν_{max}/cm^{-1} 3436 (NH), 1688 (C=O); $\delta_{\rm H}$ ([²H₆]DMSO) 10.20 (1H, s, NH), 8.34–8.72 (2H, m), 7.98 (1H, d, *J* 8.1), 7.79 (1H, t, *J* 8.0), 7.5–7.7 (3H, m), 7.26–7.29 (2H, m), 3.67 (3H, s, N–CH₃), 2.43 (2H, t, *J* 7.3, COC*H*₂), 1.60–1.72 (2H, m, CH₂), 1.32–1.47 (2H, m, CH₂CH₃), 0.95 (3H, t, *J* 7.3, CH₂CH₃); *m*/*z* (EI-HRMS) 381.1824 [C₂₅H₂₃N₃O⁺ requires 381.1841].

2-tert-Butylamido-8-methyl-8H-quino[4,3,2-kl]acridine 13d.

Method A, from 12 and pivaloyl chloride, the quinoacridine 13d was isolated as a yellow powder (89%) without the need for column chromatography. The product precipitated from the reaction mixture and was washed with DCM; mp > 250 °C; $v_{\rm max}/{\rm cm}^{-1}$ 3407 (NH), 1663 (CO); $\delta_{\rm H}$ ([²H₆]DMSO) 9.48 (1H, s, NH), 8.75 (1H, dd, *J* 1.1 and 8.0), 8.43 (1H, d, *J* 9.0), 8.36 (1H, m), 8.04 (1H, d, *J* 9.0), 7.76–7.85 (2H, m), 7.55–7.62 (2H, m), 7.23–7.31 (2H, m), 3.68 (3H, CH₃), 1.32 (9H, s, *t*-Bu); $\delta_{\rm C}$ ([²H₆]DMSO) 176.7, 149.2, 141.3, 141.2, 140.3, 133.9, 132.3, 132.0, 125.0, 123.2, 121.3, 118.5, 118.4, 117.7, 114.9, 111.0, 108.3, 45.4, 33.7, 27.2; *m*/*z* (EI-HRMS) 382.1953 [C₂₅H₂₄N₃O⁺ requires 382.1919].

2-(4-Carboxy-n-butylamido)-8-methyl-8H-quino[4,3,2-kl]-

acridine methyl ester 13e. Method A, from 12 and methyl adipoyl chloride, the quino[4,3,2-kl] acridine methyl ester was isolated as a pale yellow powder (38%), mp 202 °C (dec.);

 $v_{\rm max}/{\rm cm}^{-1}$ 3468 (NH), 1719 (C=O ester), 1686 (C=O amide); $\delta_{\rm H}([^{2}{\rm H}_{6}]{\rm DMSO})$ 10.21 (1H, s, NH), 8.77 (1H, dd, J 1.3 and 8.0), 8.45 (1H, d, J 9.0), 8.32 (1H, d, J 2.0), 8.04 (1H, d, J 8.1), 7.82 (1H, t, J 8.1), 7.56–7.66 (3H, m), 7.22–7.33 (2H, m), 3.70 (3H, s, CH_3), 3.62 (3H, s, CH_3), 2.35–2.45 (4H, m, 2 × CH_2), 1.61–1.67 (4H, m, 2 × CH_2); $\delta_{\rm C}([^{2}{\rm H}_{6}]$ DMSO) 173.2, 171.3, 149.4, 145.7, 141.2, 140.2, 133.8, 132.2, 132.0, 125.1, 123.5, 121.2, 118.3, 117.5, 116.6, 115.2, 114.9, 110.9, 108.3, 51.2, 36.1, 33.6, 33.0, 24.5, 24.1; *m/z* (EI-HRMS) 439.1894 [C₂₇H₂₅N₃O₃⁺ requires 439.1896].

2-Dodecanoylamido-8-methyl-8*H***-quino[4,3,2-***kl***]acridine 13f. Method A, from 12 and lauroyl chloride, the quinoacridine 13f was isolated as a pale yellow powder (47%), mp 225–226 °C; v_{max}/cm^{-1} 3311 (NH), 1661 (C=O); \delta_{H}[[²H₆]DMSO) 10.17 (1H, s, NH), 8.78 (1H, d,** *J* **8.4), 8.33 (1H, s), 8.05 (1H, d,** *J* **8.2), 7.83 (1H, t,** *J* **8.4), 7.57–7.65 (3H, m), 7.23–7.33 (2H, m), 3.70 (3H, s, CH₃), 2.41 (2H, t,** *J* **7.3, COCH₂), 1.66 (2H, m, COCH₂CH₂), 1.25–1.33 (16H, m, 8 × CH₂), 0.85 (3H, t,** *J* **6.8, CH₃);** *m/z* **(EI-HRMS) 479.2927 [C₃₂H₃₇N₃O⁺ requires 479.2937].**

2-Benzoylamido-8-methyl-8*H*-quino[4,3,2-*kl*]acridine 13g.

Method A, from 12 and benzoyl chloride, the benzoylamide 13g was isolated as a pale yellow powder (64%), mp > 300 °C; ν_{max}/cm^{-1} 3308 (NH), 1642 (C=O); $\delta_{\rm H}$ ([²H₆]DMSO) 8.80 (1H, d, J 7.1), 8.51–8.55 (2H, m), 8.04–8.11 (3H, m), 7.82–7.92 (2H, m), 7.55–7.66 (5H, m), 7.34 (1H, d, J 8.3), 7.27 (1H, t, J 7.0), 3.71 (3H, s, NCH₃); $\delta_{\rm H}$ ([²H₇]DMF) 10.59 (1H, s, NH), 8.89 (1H, d, J 7.5), 8.70 (1H, d, J 2.2), 8.58 (1H, d, J 9.0), 8.12–8.16 (4H, m), 7.87 (1H, t, J 8.1), 7.56–7.65 (5H, m), 7.37 (1H, d, J 8.3), 7.25–7.31 (1H, m), 3.78 (3H, s, CH₃); $\delta_{\rm C}$ [[²H₇] DMF) 166.5, 150.4, 142.2, 141.1, 135.9, 134.8, 132.8, 132.5, 132.1, 128.9, 128.3, 125.8, 123.9, 122.3, 121.7, 119.8, 119.0, 118.7, 116.3, 115.3, 111.5, 108.9; *m*/*z* (EI-HRMS) 401.1528 [C₂₇H₁₉N₃O⁺ requires 401.1528].

Synthesis of 8,13-dimethylquino[4,3,2-k/lacridinium iodides

General Method B

The appropriate substituted 8-methylacridine **13** was heated with excess methyl iodide for 2 days at 150 °C in a pressure tube. The product was collected by filtration, washed with Et_2O and purified by column chromatography (5–10% MeOH–CHCl₃). The following quinoacridinium iodides were prepared.

2-Acetylamino-8,13-dimethyl-8*H***-quino[4,3,2-***k1***]acridinium iodide 14a. Method B, from 13a (0.2 g, 0.59 mmol) and MeI (10 cm³). The acridinium iodide 14a was isolated as a bright red solid (0.25 g, 90%), mp 304 °C (dec.); v_{max}/cm^{-1} 3443 (NH), 1687 (C=O); \partial_{\rm H} ([²H₆]DMSO) 10.59 (1H, br s, NH), 8.62 (1H, d,** *J* **10.0), 8.50 (1H, d,** *J* **10.0), 8.45 (1H, d,** *J* **2.5), 8.33 (1H, d,** *J* **7.5), 8.19 (1H, t,** *J* **10.0), 8.05 (1H, t,** *J* **10.0), 8.0 (1H, d,** *J* **7.5), 7.92 (1H, d,** *J* **7.5), 7.77 (1H, dd,** *J* **2.5 and 7.5), 7.55 (1H, t,** *J* **7.5), 4.28 (3H, s, CH₃), 4.11 (3H, s, CH₃), 2.14 (3H, s, CH₃);** *m/z* **(ES-HRMS) 354.1606 [C₂₃H₂₀N₃O⁺ requires 354.1597].**

8,13-Dimethyl-2-trifluoroacetylamino-8*H***-quino[4,3,2-***kl***]-acridinium iodide 14b.** Method B, from **13b**, the iodide **14b** was isolated an orange powder (30%), mp 230 °C (dec.); v_{max} / cm⁻¹ 3445 (NH), 1719 (C=O); $\delta_{\rm H}$ ([²H₆]DMSO) 11.9 (1H, s, NH), 8.80 (1H, d, *J* 9.0), 8.56 (1H, d, *J* 8.1), 8.46–8.49 (2H, m,), 8.34 (1H, t, *J* 8.2), 8.03–8.22 (4H, m), 7.64 (1H, t, *J* 7.3), 4.37 (3H, s, CH₃), 4.20 (3H, s, CH₃); $\delta_{\rm F}$ ([²H₆]DMSO) –73.3; *m*/*z* (ES-HRMS) 408.1324 [C₂₃H₁₇F₃N₃O⁺ requires 408.1318].

2-*n***-Butylamido-8,13-dimethyl-8***H***-quino[4,3,2-***kI***]acridinium iodide 14c. Method B, from 13c, the acridinium iodide 14c was isolated as a red powder (37%), mp 166–171 °C; v_{max}/cm^{-1} 3436 (NH), 1688 (C=O); \delta_{\rm H} ([²H₆]DMSO) 8.71 (1H, d,** *J* **9.0), 8.59 (1H, d,** *J* **1.6), 8.55 (1H, d,** *J* **8.1), 8.42 (1H, d,** *J* **8.1), 8.32 (1H,** d, J 8.1), 7.94–8.20 (4H, m), 7.91 (1H, d, J 7.6), 7.61 (1H, t, J 7.6), 4.36 (3H, s, CH₃), 4.18 (3H, s, CH₃), 2.48 (2H, t, J 7.5, COCH₂), 1.62–1.73 (2H, m, COCH₂CH₂), 1.33–1.48 (2H, m, CH₂CH₃), 0.96 (3H, t, J 7.3, CH₂CH₃); $\delta_{\rm C}$ ([²H₆]DMSO) 172.5, 152.6, 143.2, 142.2, 140.0, 139.3, 136.1, 135.9, 131.5, 130.0, 125.0, 112.7, 118.6, 117.8, 117.3, 115.6, 114.8, 113.3, 112.0, 107.4, 45.9, 36.4, 36.2, 27.2, 21.8, 13.8; *m*/*z* (ES-HRMS) 396.2072 [C₂₆H₂₆N₃O⁺ requires 396.2070].

2-tert-Butylamido-8,13-dimethyl-8H-quino[4,3,2-kl]acri-

dinium iodide 14d. Method B, from **13d**, the acridinium iodide **14d** was isolated as a red powder (91%) without the need for column chromatography. Washing the product with Et₂O was sufficient to obtain pure material, mp 179 °C (dec.); $v_{\text{max}}/\text{cm}^{-1}$ 3468 (NH), 1671 (C=O); δ_{H} ([²H₆]DMSO) 9.89 (1H, s, NH), 8.70 (1H, d, J 9.1), 8.63 (1H, d, J 1.4), 8.53 (1H, d, J 8.6), 8.41 (1H, d, J 8.0), 8.29 (1H, t, J 8.0), 7.96–8.19 (4H, m), 7.62 (1H, t, J 7.3), 4.36 (3H, s, CH₃), 4.17 (3H, s, CH₃), 1.33 (9 H, s, *t*-Bu); δ_{C} ([²H₆]DMSO) 177.8, 152.8, 143.3, 142.5, 140.0, 139.2, 136.1, 135.8, 131.6, 129.8, 124.9, 122.6, 119.4, 117.9, 117.3, 115.8, 114.9, 113.4, 112.0, 108.3, 45.9, 38.5, 36.3, 27.1; *m/z* (ES-HRMS) 396.2073 [C₂₆H₂₆N₃O⁺ requires 396.2070].

2-(4-Carboxy-n-butylamido)-8,13-dimethyl-8H-quino-

[4,3,2-*kl***]acridinium iodide methyl ester 14e.** Method B, from **13e**, isolated as a red powder (40%), mp 176 °C (dec.); v_{max} /cm⁻¹ 3430 (NH), 1728 (C=O, ester), 1689 (C=O, amide); $\delta_{\rm H}$ ([²H₆]DMSO) 10.59 (1H, s, NH), 8.46–8.56 (3H, m), 8.06–8.28 (4H, m,), 7.91 (1H, d, *J* 8.2), 7.78 (1H, d, *J* 8.8), 7.60 (1H, t, *J* 7.2), 4.30 (3H, s, CH₃), 4.13 (3H, s, CH₃), 3.62 (3H, s, CH₃), 2.5 (2H, m, CH₂), 2.41 (2H, t, *J* 7.0, CH₂), 1.62–1.70 (4H, m, 2 × CH₂); $\delta_{\rm C}$ ([²H₆]DMSO) 173.2, 172.0, 152.5, 143.1, 142.1, 139.9, 139.1, 135.9, 135.8, 131.3, 129.6, 124.9, 122.6, 118.4, 117.7, 117.2, 115.5, 114.7, 113.2, 111.9, 107.2, 51.3, 45.9, 36.2, 33.1, 24.4, 24.1; *m*/*z* (ES-HRMS) 454.2146 [C₂₈H₂₈N₃O₃⁺ requires 454.2131].

2-Dodecanoylamido-8,13-dimethyl-8H-quino[4,3,2-kl]-

acridinium iodide 14f. Method B, from **13f**, isolated as a red powder (45%), mp 182–184 °C; v_{max}/cm^{-1} 3435 (NH), 1689 (CO); $\delta_{\rm H}([^{2}{\rm H}_{6}]{\rm DMSO})$ 10.55 (1H, s, NH), 8.44–8.52 (3H, m), 8.02–8.24 (4H, m), 7.89 (1H, d, *J* 8.0), 7.78 (1H, d, *J* 8.7), 7.60 (1H, t, *J* 7.8), 4.29 (3H, s, CH₃), 4.11 (3H, s, CH₃), 2.45 (2H, t, *J* 7.3, COCH₂), 1.67 (2H, m, COCH₂CH₂), 1.25–1.33 (16H, m, 8 × CH₂), 0.86 (3H, t, *J* 6.6, CH₃), $\delta_{\rm c}([^{2}{\rm H}_{6}]{\rm DMSO})$ 172.4, 152.6, 143.2, 142.2, 140.0, 139.3, 136.0, 135.8, 131.5, 125.0, 122.6, 118.5, 117.7, 117.3, 115.6, 114.9, 113.3, 112.0, 107.2, 45.9, 36.6, 36.2, 29.1, 29.0, 29.0, 28.9, 28.8, 28.7, 31.3, 22.1, 14.0; *m/z* (ES-HRMS) 494.3131 [C₃₃H₄₀N₃O⁺ requires 494.3171].

2-Benzoylamido-8,13-dimethyl-8*H***-quino[4,3,2-***kI***]acridinium iodide 14g. Method B, from 13g, isolated as a red powder (18%), mp 180 °C (dec.); v_{max}/cm⁻¹ 3429 (NH), 1670 (C=O); \delta_{\rm H} ([²H₆]DMSO) 10.86 (1H, s, NH), 8.67 (2H, m), 8.56 (1H, d,** *J* **8.1), 7.94–8.38 (8H, m), 7.60–7.70 (4H, m), 4.37 (3H, s, CH₃), 4.15 (3H, s, CH₃); \delta_{\rm C} ([²H₆]DMSO) 166.2, 152.6, 143.2, 142.1, 140.0, 139.1, 136.0, 135.8, 134.3, 132.2, 131.4, 129.8, 128.6, 127.8, 124.9, 122.7, 119.5, 118.2, 117.3, 115.7, 114.8, 113.4, 112.1, 108.6, 46.0, 36.2;** *m***/***z* **(ES-HRMS) 416.1765 [C₂₈H₂₂-N₃O⁺ requires 416.1763].**

2-Iodo-8-methyl-8*H*-quino[4,3,2-*kl*]acridine 16

A solution of sodium nitrite (87 mg, 1.26 mmol) in water (2 cm^3) was added dropwise to a solution of 2-amino-8-methyl-8*H*-quino[4,3,2-*kl*]acridine (250 mg, 0.84 mmol) in 2 M HCl (50 cm³) at 0 °C. After stirring at 0 °C for 15 min, a solution of KI (0.24 g, 1.47 mmol) in water (10 cm³) was added dropwise and the resulting mixture heated for 0.5 h at 80 °C. After cooling, the reaction mixture was basified (to pH 12) with 10% sodium hydroxide and the product extracted with CHCl₃ (4 × 50 cm³). Purification by column chromatography (0.25% MeOH–CHCl₃) gave **16** as a yellow solid (190 mg, 55%), mp 265–268 °C (Found: C, 58.41; H, 3.27; N, 6.44. C₂₀H₁₃N₂I requires C, 58.82; H, 3.19; N, 6.86%); v_{max}/cm^{-1} 1597 (C=N); $\delta_{\rm H}$ ([²H₆]DMSO) 8.78 (1H, dd, *J* 1.5 and 9.3), 8.32 (1H, d, *J* 8.8), 8.27 (1H, d, *J* 2.0), 8.13 (1H, d, *J* 7.8), 7.89 (1H, t, *J* 8.0), 7.77 (1H, dd, *J* 1.8 and 8.5), 7.66–7.63 (2H, m), 7.44 (1H, d, *J* 8.3), 7.29 (1H, t, *J* 8.0), 3.73 (3H, s, CH₃); MS (ES) *m/z* 409.2 (M + 1); *m/z* (ES-HRMS) 409.0214 [C₂₀H₁₄N₂I⁺ requires 409.0202].

8,13-Dimethyl-2-iodo-8*H*-quino[4,3,2-*kl*]acridinium iodide 17

Method B, from **16** (57 mg, 0.14 mmol) furnished **17** as a bright red solid (65 mg, 85%), mp 240–243 °C; ν_{max}/cm^{-1} 1611 (C=N); $\delta_{\rm H}$ ([²H₆]DMSO) 8.55–8.45 (4H, m), 8.34 (1H, t, *J* 8.25), 8.23–8.06 (4H, m), 7.65 (1H, t, *J* 7.25), 4.36 (3H, s, CH₃), 4.21 (3H, s, CH₃); MS (ES) *m*/*z* 423.0 (M⁺).

2-*tert*-Butoxycarbonylamino-8-methyl-8*H*-quino[4,3,2-*kl*]-acridine 18

Di-*tert*-butyl dicarbonate (Boc anhydride; 0.8 g, 3.7 mmol) was added to a solution 2-amino-8-methyl-8*H*-quino-[4,3,2-*kl*]-acridine (1 g, 3.36 mmol) in THF (20 cm³). A 10% solution of NaHCO₃ (10 cm³) was added and the resulting mixture heated under reflux for 3 h. After cooling, CHCl₃ (50 cm³) was added, the organic phase removed and washed with water (10 cm³). The organic phase was purified by passing through a short bed of silica, followed by washing with 3% MeOH–DCM to leave **18** as a yellow solid (1.3 g, 97%), mp 245–248 °C; $\nu_{\rm max}/\rm cm^{-1}$ 3213 (NH), 1720 (C=O); $\delta_{\rm H}$ ([²H₆]DMSO) 9.71 (1H, br s, NH), 8.76 (1H, d, *J* 7.75), 8.42 (1H, d, *J* 9.0), 8.11 (1H, d, *J* 2.25), 8.02 (1H, d, *J* 8.0), 7.75 (1H, t, *J* 8.0), 7.63–7.56 (3H, m), 7.31–7.22 (2H, m), 3.7 (3H, s, NCH₃), 1.56 (9H, s, $3 \times \rm CH_3$); *m/z* (ES-HRMS) 398.1882 [C₂₅H₂₄N₃O₂⁺ requires 398.1869).

2-*tert*-Butoxycarbonylamino-8,13-dimethyl-8*H*-quino[4,3,2-*kl*]-acridinium iodide 19

By Method B, from 2-*tert*-butoxycarbonylamino-8-methyl-8*H*-quino[4,3,2-*kl*]acridine (0.2 g, 0.5 mmol) and methyl iodide (10 cm³) for 7 days at 80 °C in a sealed tube, the crude quaternary salt was purified by column chromatography (10% MeOH–CHCl₃) to afford **19** as a bright red solid (200 mg, 73%), mp 183–185 °C; v_{max}/cm^{-1} 3424 (NH), 1724 (C=O); $\delta_{\rm H}$ ([²H₆]DMSO) 10.16 (1H, br s, NH), 8.65 (1H, d, *J* 10.0), 8.55 (1H, d, *J* 10.0), 8.39–8.25 (3H, m), 8.16 (1H, d, *J* 7.5), 8.08 (1H, dd, *J* 2.5 and 10.0), 7.96 (1H, d, *J* 7.5), 7.62 (1H, d, *J* 7.5), 7.60 (1H, t, *J* 7.5), 4.35 (3H, s, CH₃), 4.16 (3H, s, CH₃), 1.58 (9H, s, 3 × CH₃); *m/z* (ES-HRMS) 412.2002 [C₂₆H₂₆N₃O₂⁺ requires 412.2025].

2-Amino-8,13-dimethyl-8*H*-quino[4,3,2-*k1*]acridinium chloride 20a

2-*tert*-Butoxycarbonylamino-8,13-dimethyl-8*H*-quino[4,3,2-*kl*]-acridinium iodide **19** or 2-acetylamino-8,13-dimethyl-8*H*-quino-[4,3,2-*kl*]acridinium iodide **14a** (0.3 mmol) were dissolved in a mixture of methanol (50 cm³) and 2 M HCl (50 cm³). The resulting solution was stirred at 50 °C for 2 h. The methanol was removed under vacuum and the aqueous phase basified (to pH 10) with 5 M NaOH. The resulting precipitate was filtered from solution and washed with water to yield **20a** as a dark purple solid (92%), mp 215–218 °C; v_{max} /cm⁻¹ 3410 (NH), 1608 (C=N); $\delta_{\rm H}$ ([²H₆]DMSO) 8.47 (1H, d, *J* 8.5), 8.40 (1H, d, *J* 8.8), 8.21–8.05 (4H, m), 7.75–7.67 (2H, m), 7.55 (1H, t, *J* 7.8), 7.07 (1H, d, *J* 2.5), 7.04 (1H, dd, *J* 2.5 and 8.8), 6.44 (2H, br s, NH₂), 4.28 (3H, s, CH₃), 4.08 (3H, s, CH₃); *m/z* (ES-HRMS) 312.1499 [C₂₃H₂₂N₃⁺ requires 312.1495].

Methylation of 2-*tert*-butoxycarbonylamino-8-methyl-8*H*-quino-[4,3,2-*kl*]acridine 18

2-*tert*-Butoxycarbonylamino-8-methyl-8*H*-quino[4,3,2-*kl*]acridine **18** (0.5 g, 1.26 mmol) was suspended in iodomethane (10 cm³) and heated for 48 h at 150 °C in a sealed tube. After cooling, the precipitate was filtered from solution and purified by column chromatography (5% MeOH–CHCl₃) leading to the isolation of 3 dark purple solids.

2-Amino-8,13-dimethyl-8*H***-quino[4,3,2-***kI***]acridinium iodide 20b.** (30 mg, 5%), Mp 222–225 °C; v_{max}/cm^{-1} 3400 (NH), 1611 (C=N); $\delta_{\rm H}$ ([²H₆]DMSO) 8.47 (1H, d, *J* 8.5), 8.40 (1H, d, *J* 8.75), 8.21–8.02 (4H, m), 7.73 (1H, m), 7.55 (1H, t, *J* 7.75), 7.07 (1H, d, *J* 2.25), 7.04 (1H, dd, *J* 2.25 and 8.5), 6.44 (2H, br s, NH₂), 4.28 (3H, s, CH₃), 4.08 (3H, s, CH₃); *m*/*z* (APCI-HRMS) 312.1482 [C₂₁H₁₈N₃⁺ requires 312.1501].

8,13-Dimethyl-2-methylamino-8*H*-quino[**4,3,2**-*kI*]acridinium iodide **20c.** (100 mg, 17%), Mp 156–158 °C; v_{max} /cm⁻¹ 3239 (NH), 1617 (C=N); $\delta_{\rm H}$ ([²H₆]DMSO) 8.42 (2H, m), 8.14–7.99 (4H, m), 7.70 (1H, dd, *J* 2.0 and 7.0), 7.56 (1H, t, *J* 7.5), 7.04 (2H, m), 6.85 (1H, br s, NH), 4.34 (3H, s, CH₃), 4.03 (3H, s, CH₃), 2.94 (3H, d, *J* 4.75, CH₃); *m/z* (ES-HRMS) 326.1642 [C₂₂H₂₀N₃⁺ requires 326.1657].

8,13-Dimethyl-2-dimethylamino-8*H***-quino[4,3,2-***kI***]acridinium iodide 20d. (250 mg, 63%), Mp 220–222 °C; v_{max}/cm⁻¹ 1621, 1607 (C=N); \delta_{\rm H} ([²H₆]DMSO) 8.46 (1H, d,** *J* **9.5), 8.43 (1H, d,** *J* **9.5), 8.22–8.07 (4H, m), 7.73 (1H, d,** *J* **8.0), 7.60 (1H, t,** *J* **7.75), 7.21 (1H, dd,** *J* **2.25 and 9.0), 6.94 (1H, d,** *J* **2.25), 4.37 (3H, s, CH₃), 4.16 (3H, s, CH₃);** *m/z* **(ES-HRMS) 340.1817 [C₂₃H₂₂N₃⁺ requires 340.1814].**

3-Benzylamino-8-methyl-8H-quino[4,3,2-kl]acridine 25a

3-Chloro-8-methyl-8*H*-quino[4,3,2-kl] acridine 23⁷ (100 mg, 032 mmol), benzylamine (50 mg, 0.47 mmol) and Pd₂(dba)₃ (6 mg, 0.0063 mmol) were dissolved in dioxane (10 cm³) and placed in a sealed tube under nitrogen. Sodium tert-butoxide (60 mg, 0.64 mmol) and tri-tert-butylphosphine (2.5 mg, 0.012 mmol) were added and the resulting solution heated for 24 h at 120 °C. After cooling, the reaction mixture was filtered through a short bed of Celite, which was subsequently washed with DCM (50 cm³). Concentration of the organic phase, followed by column chromatography (25% EtOAc-hexane) gave 25a as a dark red solid (0.12 g, 96%), mp 186-188 °C; v_{max}/cm^{-1} 3416 (NH), 1620 (C=N); δ_{H} ([²H₆]DMSO) 8.65 (1H, dd, J 1.5, 8.0), 7.68-7.81 (3H, m), 7.52-7.14 (11H, m), 6.68 (1H, t, J 6.0, NH), 4.50 (2H, d, J 6.0, CH₂), 3.61 (3H, s, CH₃); *m/z* (APCI-HRMS) 388.1807 [C₂₇H₂₂N₃⁺ requires 388.1808].

3-Amino-8-methyl-8H-quino[4,3,2-kl]acridine 25b

3-Benzylamino-8-methyl-8*H*-quino[4,3,2-*kl*]acridine (50 mg, 0.13 mmol) and hydrazine hydrate (40 mg, 0.6 mmol) were added to a suspension of 10% palladium on carbon (50 mg) in methanol (10 cm³) and heated under reflux for 4 h. After cooling, the catalyst was removed by filtration and the filtrate concentrated to dryness. The residue was purified by column chromatography (5% MeOH–CHCl₃) to leave **25b** as a dark red solid (34 mg, 88%), mp > 250 °C; v_{max}/cm^{-1} 3423 (NH), 1606 (C=N); $\delta_{\rm H}$ ([²H₆]DMSO) 8.50 (1H, dd, *J* 1.5, 8.0), 7.67–7.58 (2H, m), 7.53 (1H, d, *J* 8.75), 7.48–7.32 (3H, m), 7.13–6.98 (2H, m), 6.92 (1H, dd, *J* 2.25, 8.75), 5.40 (2H, br s, NH₂), 3.49 (3H, s, CH₃); $\delta_{\rm C}$ ([²H₆]DMSO) 147.6, 145.0, 142.2, 141.7, 138.2, 133.5, 132.4, 131.8, 130.4, 125.2, 124.6, 122.7, 121.9, 120.1, 116.7, 115.4, 111.8, 109.2, 104.0, 34.4; *m/z* (APCI-HRMS) 298.1344 [C₂₀H₁₆N₃⁺ requires 298.1339].

3-Acetamido-8-methyl-8H-quino[4,3,2-kl]acridine 25c

Acetic anhydride (0.18 g, 1.7 mmol) was added dropwise to a solution of 3-amino-8-methyl-8*H*-quino[4,3,2-*kl*]acridine (0.26 g, 0.87 mmol) in pyridine (5 cm³). The resulting solution was heated for 2 h at 100 °C, cooled and poured into water (20 cm³). The product was extracted with CHCl₃ (5 × 10 cm³) and the extracts washed with water (10 cm³), dried (Na₂SO₄), and evaporated. The residue was purified by column chromatography (5% MeOH–CHCl₃) to furnish **25c** as a dark red solid (0.2 g, 67%), mp > 300 °C (dec.). v_{max} /cm⁻¹ 3272 (NH), 1659 (C=O); $\delta_{\rm H}$ ([²H₆]DMSO) 10.27 (1H, br s, NH), 8.84 (1H, d, *J* 1.75), 8.76 (1H, dd, *J* 1.25, 7.75), 7.90–7.81 (4H, m), 7.78–7.56 (2H, m), 7.37 (1H, d, 7.75), 7.26 (1H, d, *J* 7.75), 3.70 (3H, s, NCH₃), 2.15 (3H, s, CH₃); *m*/*z* (APCI-HRMS) 340.1444 [C₂₂H₁₈N₃O⁺ requires 340.1444].

8-Methyl-3-(morpholin-4-yl)-8*H*-quino[4,3,2-*kl*]acridine 25d

3-Chloro-8-methyl-8*H*-quino[4,3,2-*kl*]acridine **23**⁷ (50 mg, 0.16 mmol), morpholine (28 mg, 2 mol eq.), Pd(OAc)₂ (2 mg, 5 mol%), P(*t*-Bu)₃ (0.32 cm³, 0.05 M in dioxane 10 mol%), sodium *tert*-butoxide (22 mg, 1.4 mol eq.) and dioxane (1 cm³) were heated under nitrogen in a screw-top reaction vessel at 150 °C for 72 h. Products were dissolved in DCM, purified by column chromatography (5% MeOH–DCM) and crystallised from CHCl₃ to furnish the 3-morpholinyl-quinoacridine **25d** as an orange solid (28 mg, 48%), mp 259–261 °C; v_{max} / cm⁻¹ 1610 (C=N); $\delta_{\rm H}$ ([²H₆]DMSO) 8.72 (1H, d, *J* 7.5, H-12), 8.13 (1H, d, *J* 8.0, H-5), 7.85–7.81 (3H, m, H-1, H-4, H-6), 7.58–7.51 (3H, m, H-2, H-9, H-10), 7.31 (1H, d, *J* 8.5, H-7), 7.22 (1H, d, *J* 7.0, H-11), 4.14 (2H, m, CH₂), 3.84 (6H, br s, 3 × CH₂), 3.68 (3H, s, NCH₃); *m/z* (EI-HRMS) 367.1685 [C₂₄H₂₁N₃O⁺ requires 367.1685].

3-Cyano-8-methyl-8H-quino[4,3,2-kl]acridine 25e

The 3-chloroquinoacridine **23**⁷ (0.15 g, 0.47 mmol), Pd₂(dba)₃ (9 mg), dppf (17 mg), Zn powder (5 mg), Zn(CN)₂ (33 mg) and dimethylacetamide (20 cm³) were heated in a pressure tube under nitrogen at 100 °C for 72 h. Products were extracted into DCM (3 × 20 cm³) and washed with 2 M aqueous ammonia (20 cm³) and water (20 cm³). The organic layer was filtered, concentrated under reduced pressure and fractionated by column chromatography (2% MeOH–CHCl₃) to give the cyanoquinoacridine **25e** (0.147 g, 99%), mp 280 °C (dec.); v_{max}/cm^{-1} 2215 (CN), 1591; $\delta_{\rm H}$ ([²H₆]DMSO) 9.11 (1H, s, H-4), 8.81 (1H, d, *J* 8.0, H-12), 8.30 (1H, d, *J* 7.5, H-7), 7.98–7.92 (3H, m, H-1, H-2, H-6), 7.72–7.66 (2H, m, H-9, H-10), 7.54 (1H, d, *J* 7.5, H-7), 7.32 (1H, t, *J* 8.0, H-11), 3.77 (3H, s, 8-Me); *m/z* (EI-HRMS) 307.1096 [C₂₁H₁₃N₃ requires 307.1110].

3-Acetamido-8,13-dimethyl-8*H*-quino[4,3,2-*kl*]acridinium iodide 26

3-Acetamido-8-methyl-8*H*-quino[4,3,2-*kl*]acridine (0.2 g, 0.5 mmol) was suspended in methyl iodide (10 cm³) and heated in a sealed tube for 3 days at 150 °C. After cooling, Et₂O (40 cm³) was added and the precipitate collected as a bright red solid (0.2 g, 70%), mp 231–233 °C; v_{max}/cm^{-1} 3436 (NH), 1685 (C=O); $\delta_{\rm H}$ ([²H₆]DMSO) 10.52 (1H, br s, NH), 9.00 (1H, d, *J* 6.5), 8.48 (1H, d, *J* 8.25), 8.35 (1H, t, *J* 8.25), 8.18–7.97 (6H, m), 7.61 (1H, t, 8.25), 4.39 (3H, s, CH₃), 4.16 (3H, s, CH₃), 2.19 (3H, s, CH₃); *m/z* (ES-HRMS) 354.1607 [C₂₃H₂₀N₃O⁺ requires 354.1597].

Acknowledgements

This work was supported by Cancer Research UK through a studentship (to R. A. H) and project and programme support to the Cancer Research UK Experimental Cancer Chemotherapy Research Group.

References

- 1 R. A. Heald and M. F. G. Stevens, Org. Biomol. Chem., 2003, 1, 3377–3389.
- 2 G. Mitchell and C. W. Rees, J. Chem. Soc., Perkin Trans. 1, 1987, 403-412.
- 3 C. Graebe and F. Ullmann, *Justus Liebigs Ann. Chem.*, 1896, **291**, 16–24.
- 4 D. J. Hagan, E. Giménez-Arnau, C. H. Schwalbe and M. F. G. Stevens, J. Chem. Soc., Perkin Trans. 1, 1997, 2739–2746.
- 5 M. J. Ellis and M. F. G. Stevens, J. Chem. Soc., Perkin Trans. 1, 2001, 3180–3185.
- 6 M. F. G. Stevens, L. R. Kelland and R. A. Heald, PCT/GB01/ 04557.
- 7 R. A. Heald, C. Modi, J. C. Cookson, I. Hutchinson, C. A.

Laughton, S. M. Gowan, L. R. Kelland and M. F. G. Stevens, J. Med. Chem., 2002, 45, 590–597.

- 8 A. Hassner and C. Stumer, in Organic Reactions Based on Name Reactions and Unnamed Reactions, Elsevier Science Ltd., Oxford, 1994, p. 307; P. H. Leake, Chem. Rev., 1956, 56, 27–48.
- 9 A. R. Katritzky, W. Du, Y. Matsukawa, I. Ghiviriga and S. N. Denisenko, J. Heterocyclic Chem., 1999, **36**, 927–932.
- 10 F. Jin and P. N. Confalone, Tetrahedron Lett., 2000, 41, 3271-3273.
- 11 M. R. Boyd and K. D. Paull, Drug Dev. Res., 1995, 34, 91-104.
- 12 J. N. Weinstein, T. G. Myers, P. M. O'Connor, S. H. Friend, A. J. Fornace, K. W. Kohn, T. Fojo, S. E. Bates, L. V. Rubenstein, N. L. Anderson, J. K. Buolamwini, W. W. van Osdol, A. P. Monks, D. A. Scudiero, E. A. Sausville, D. W. Zaharevitz, B. Bunow, V. N. Viswanadhan, G. S. Johnson, R. E. Wittes and K. D. Paull, *Science*, 1997, **275**, 343–349.